

Kinetics of protein precipitation: optimizing recovery, purity, and throughput using the ProTrap XG

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OBJECTIVES



- Optimize the efficiency of protein precipitation
- Determine the versatility of precipitation for dilute samples
- Quantify SDS depletion; determine whether MS threshold of <10 ppm is achieved
- Determine digestion efficiency within the ProTrap XG
- Analyze proteome coverage, determining whether precipitation favors proteins of high MW, or particular pI or GRAVY score

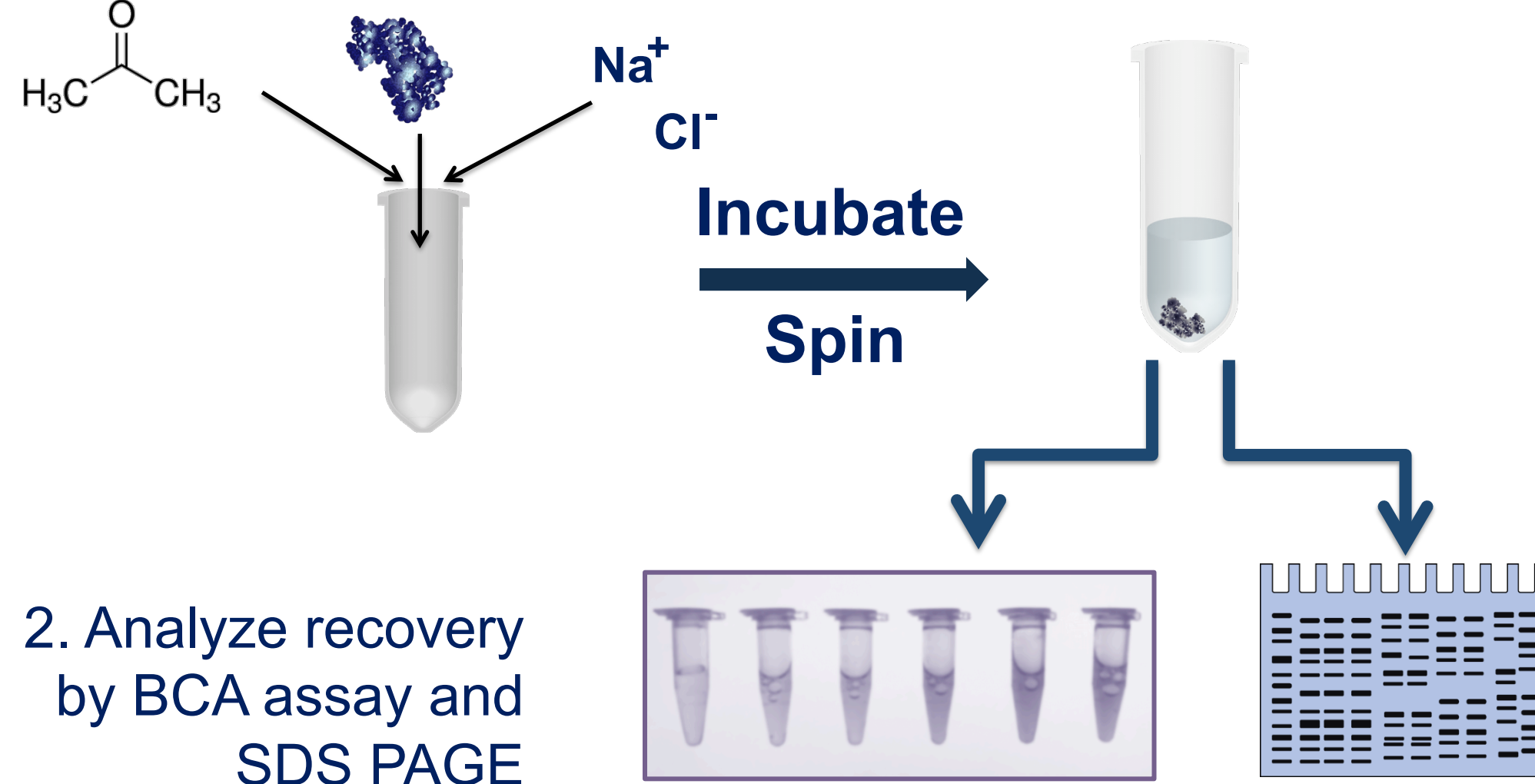
INTRODUCTION

Efficient sample preparation is key to any proteomics workflow, with three priorities at the forefront: protein recovery, purity, and sample throughput. Acetone precipitation is a common method of purifying proteins from a complex sample prior to LC-MS/MS analysis, but is sometimes set aside due to seemingly variable yields.¹ Our group previously established that the addition of 1-30 mM salt with 80% v/v acetone facilitates recoveries >95%.² In order to maximize recovery, purity and sample throughput, we investigated the kinetics of protein precipitation with respect to structural properties of proteins, including molecular weight, hydrophobicity, and charge.

METHODS

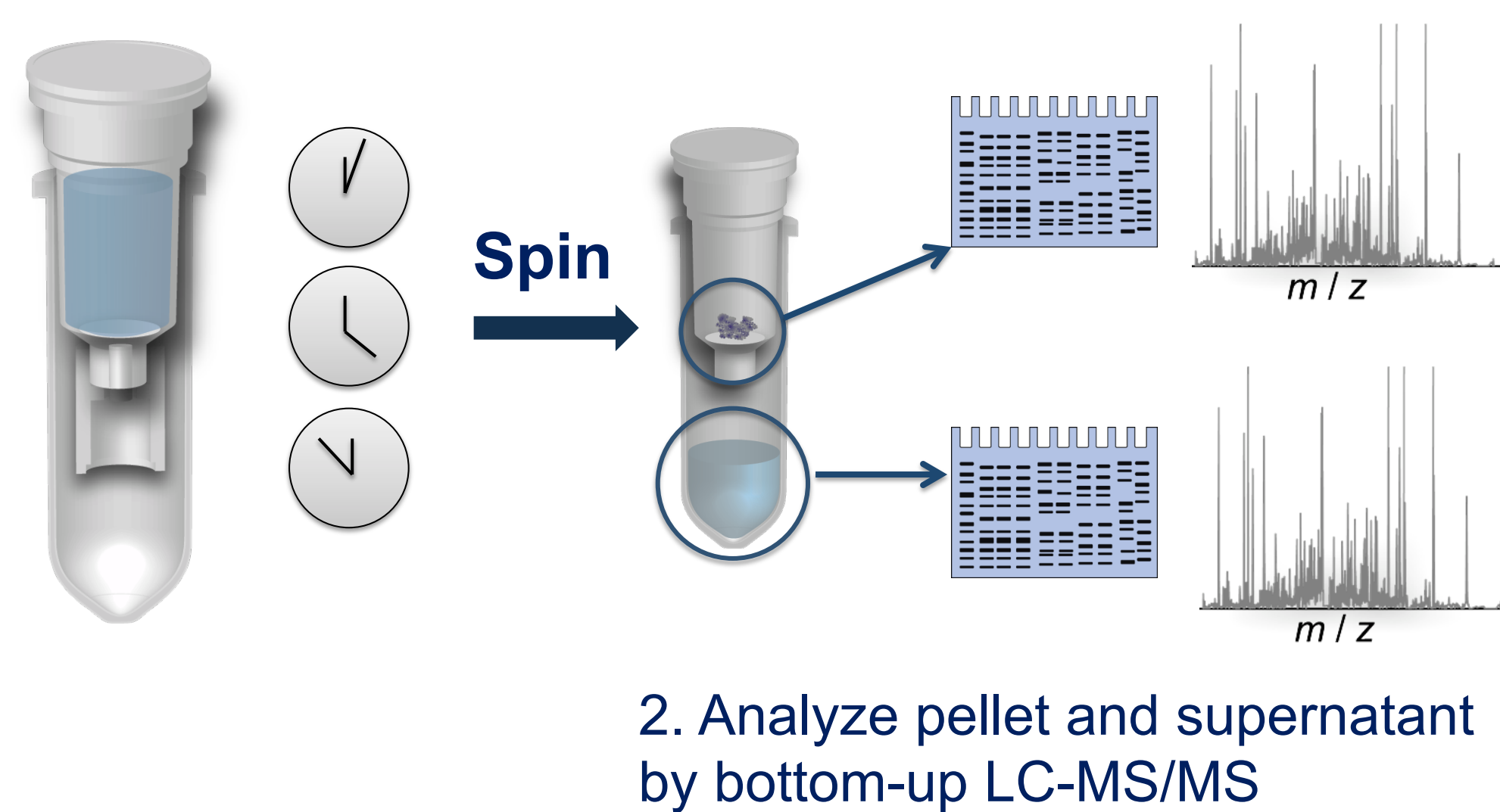
Optimize precipitation protocol

- Precipitate protein with 0.1 - 100 mM NaCl and 80 % acetone

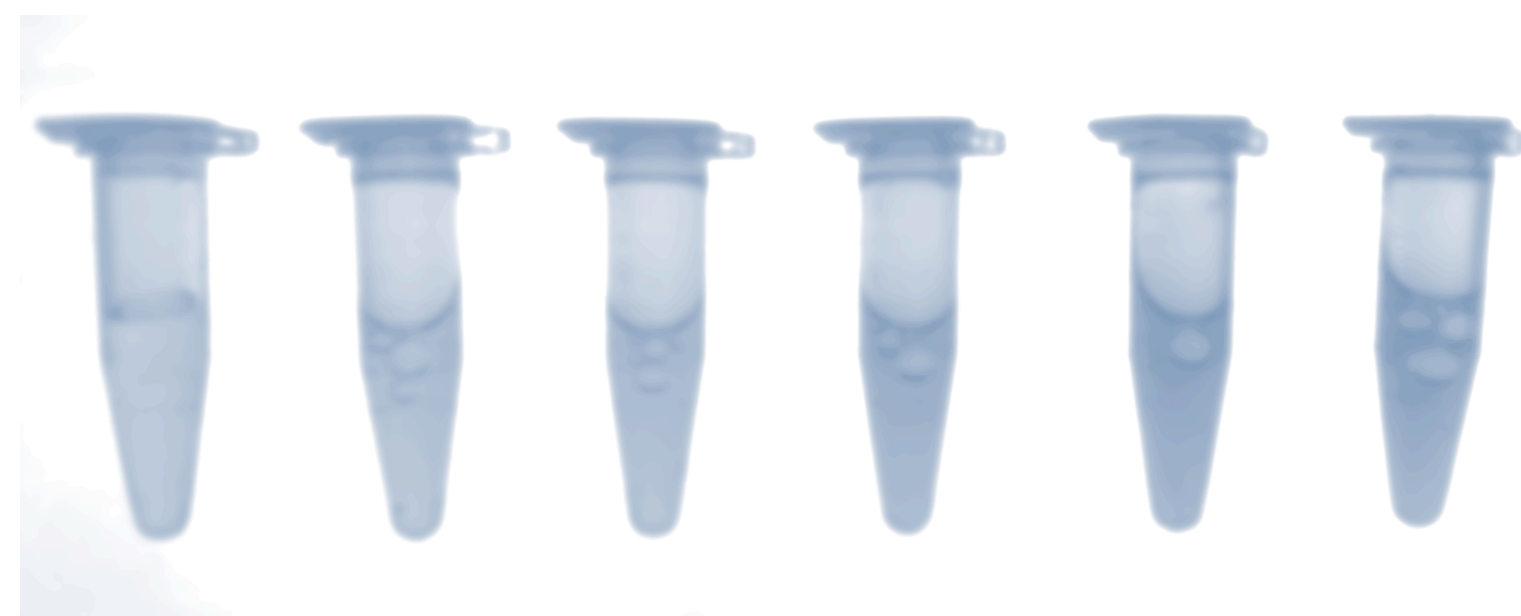
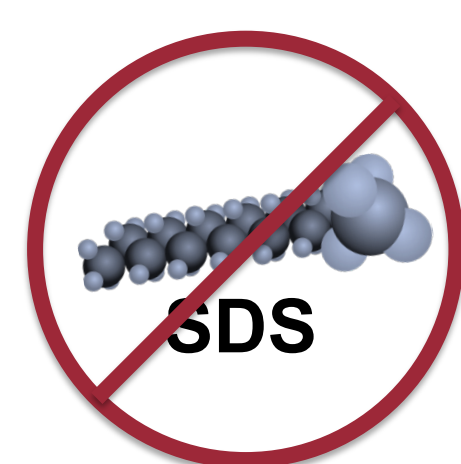


Precipitation in the ProTrap XG

- Precipitate yeast proteome with 20 mM NaCl and 4 volumes acetone

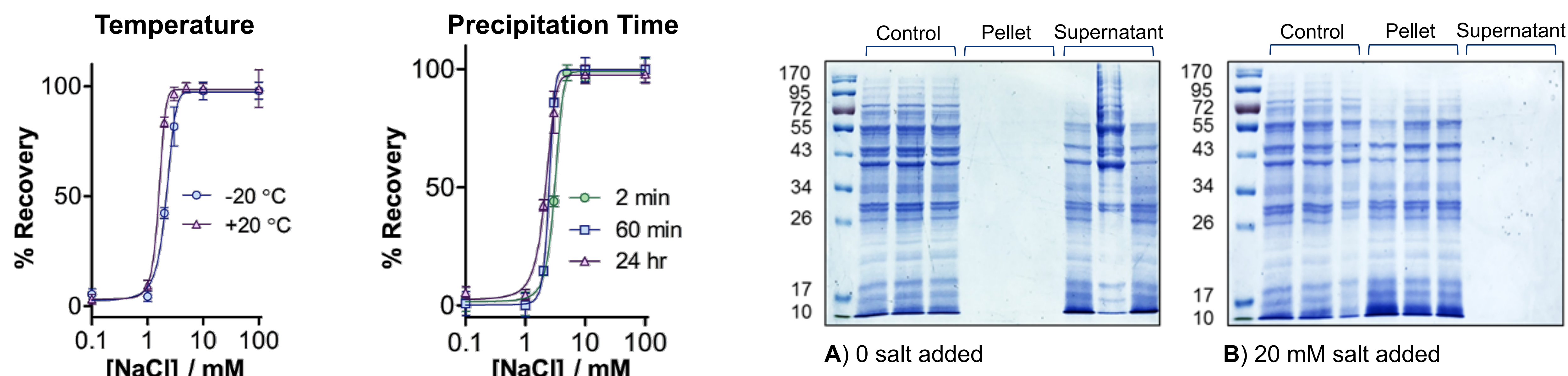


- Determine purity by quantifying residual SDS by MBAS assay

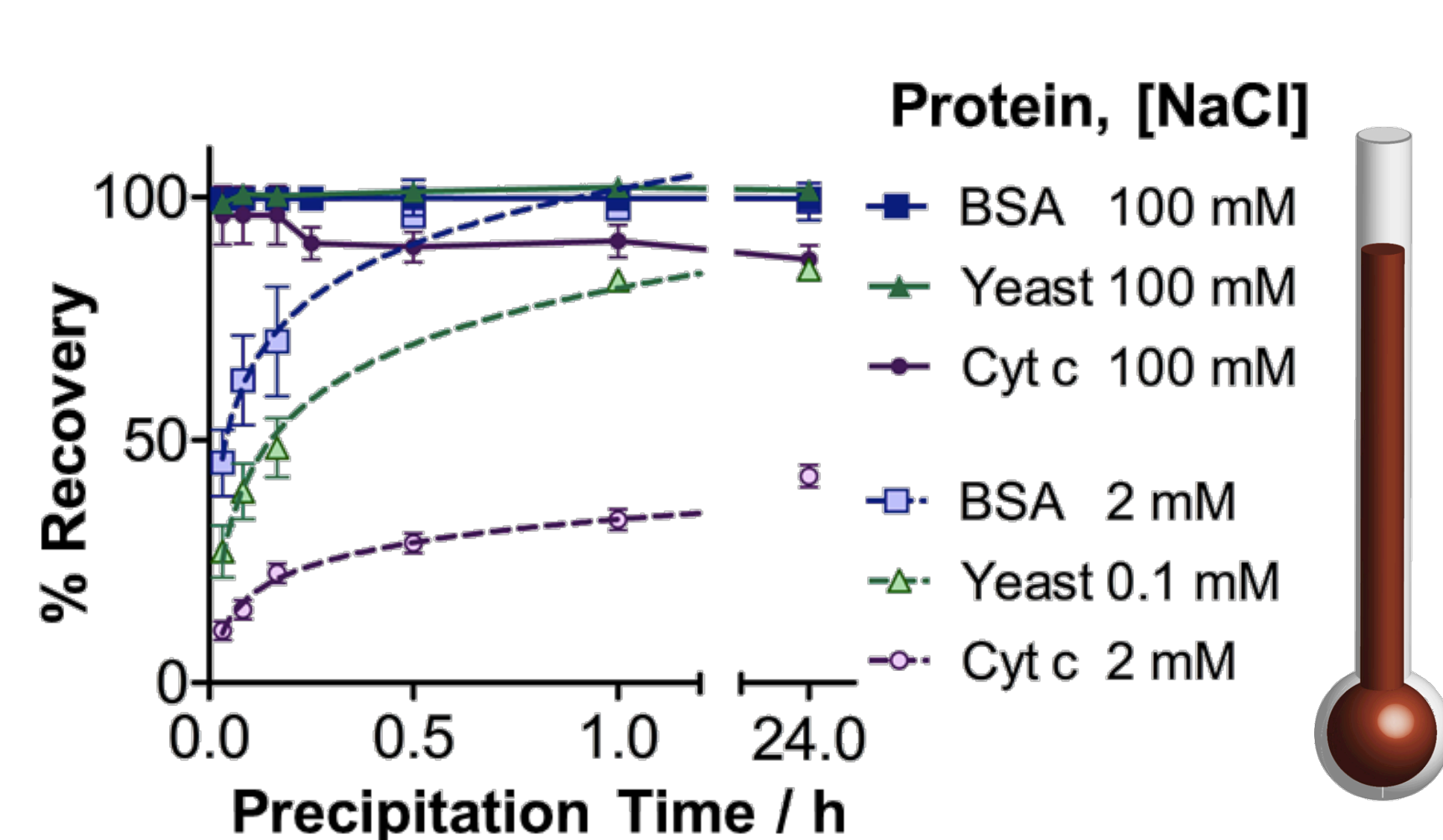


RESULTS

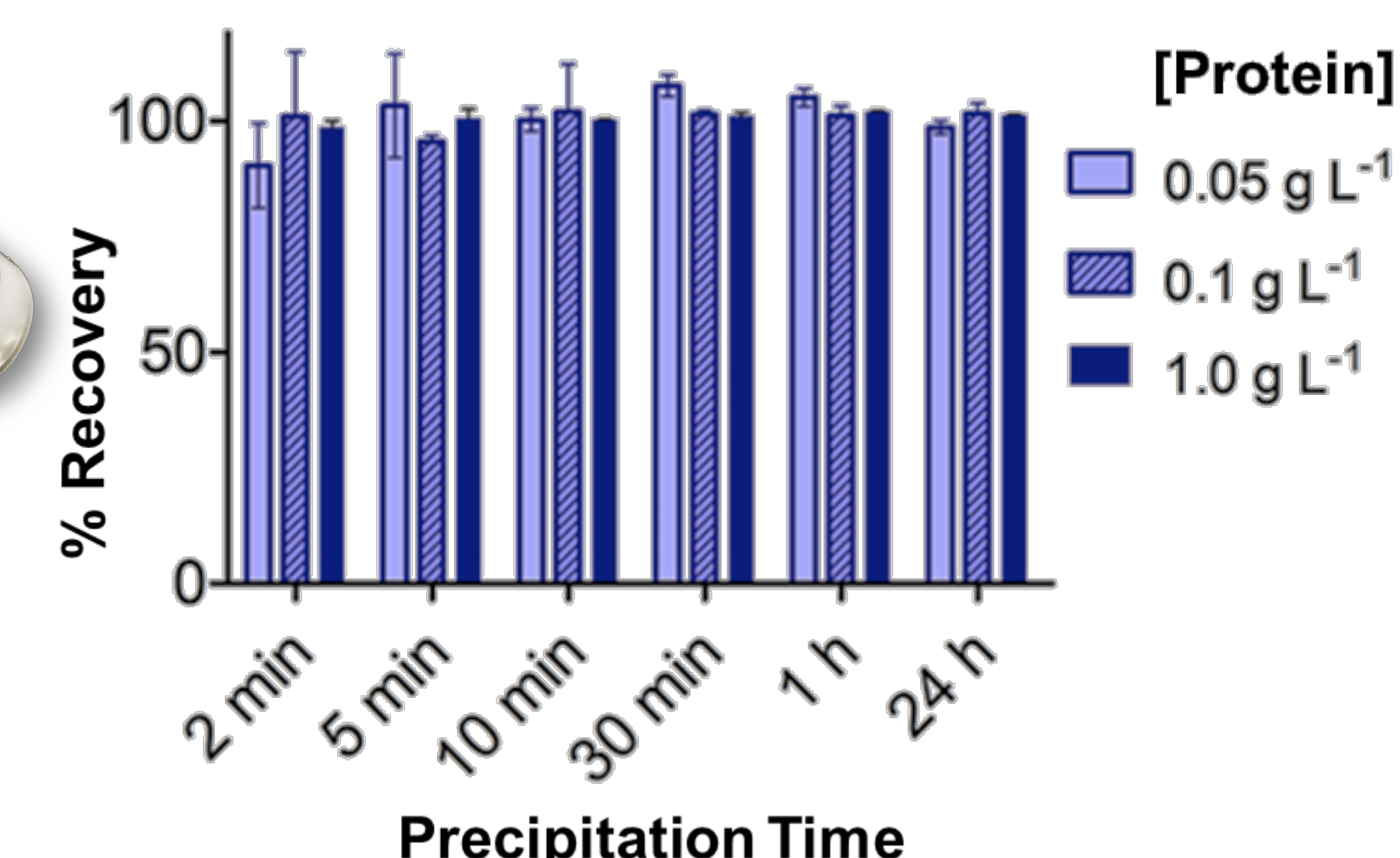
High salt is critical for high recovery in 2 min



High salt improves precipitation kinetics

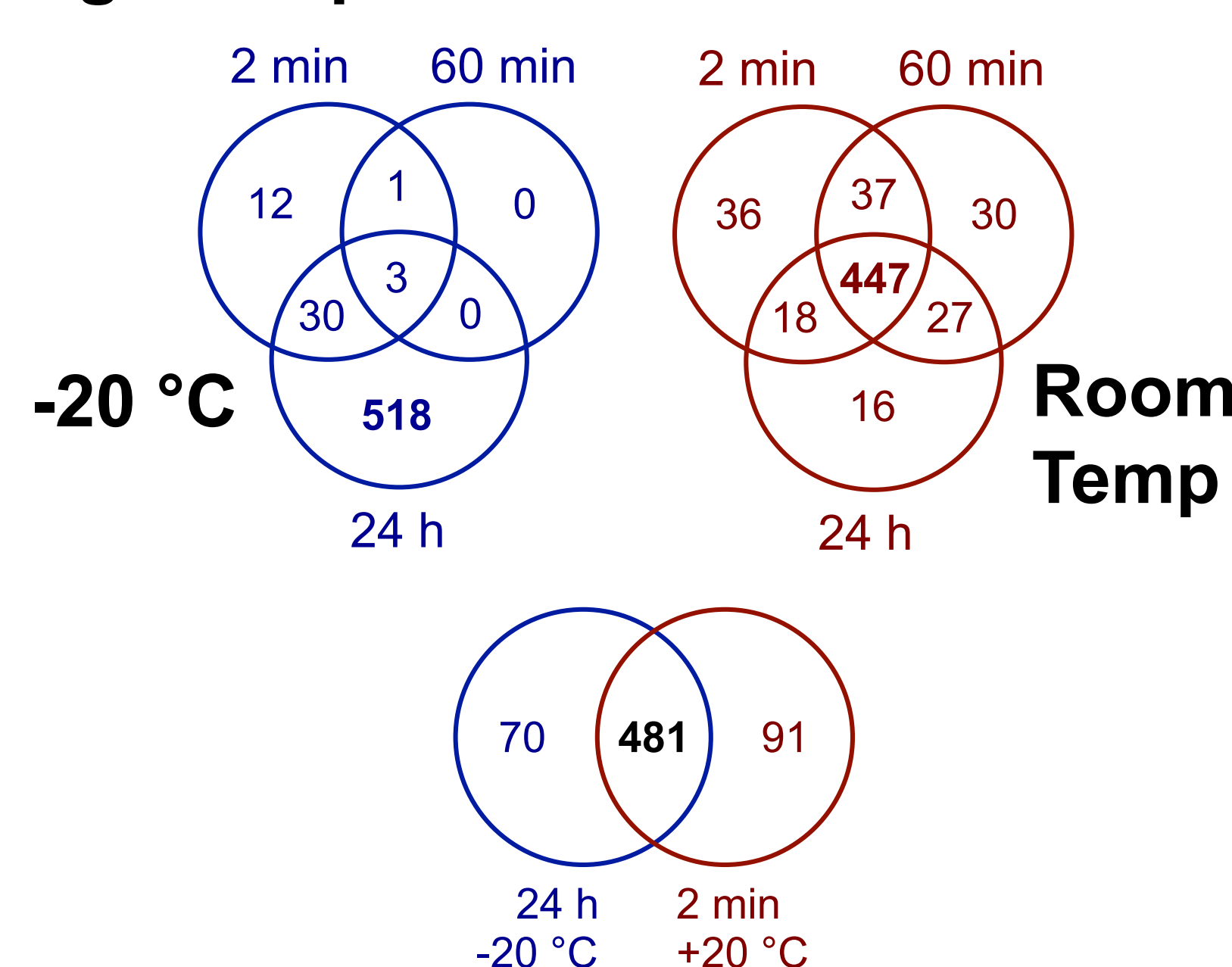


Dilute samples precipitate in 2-5 min

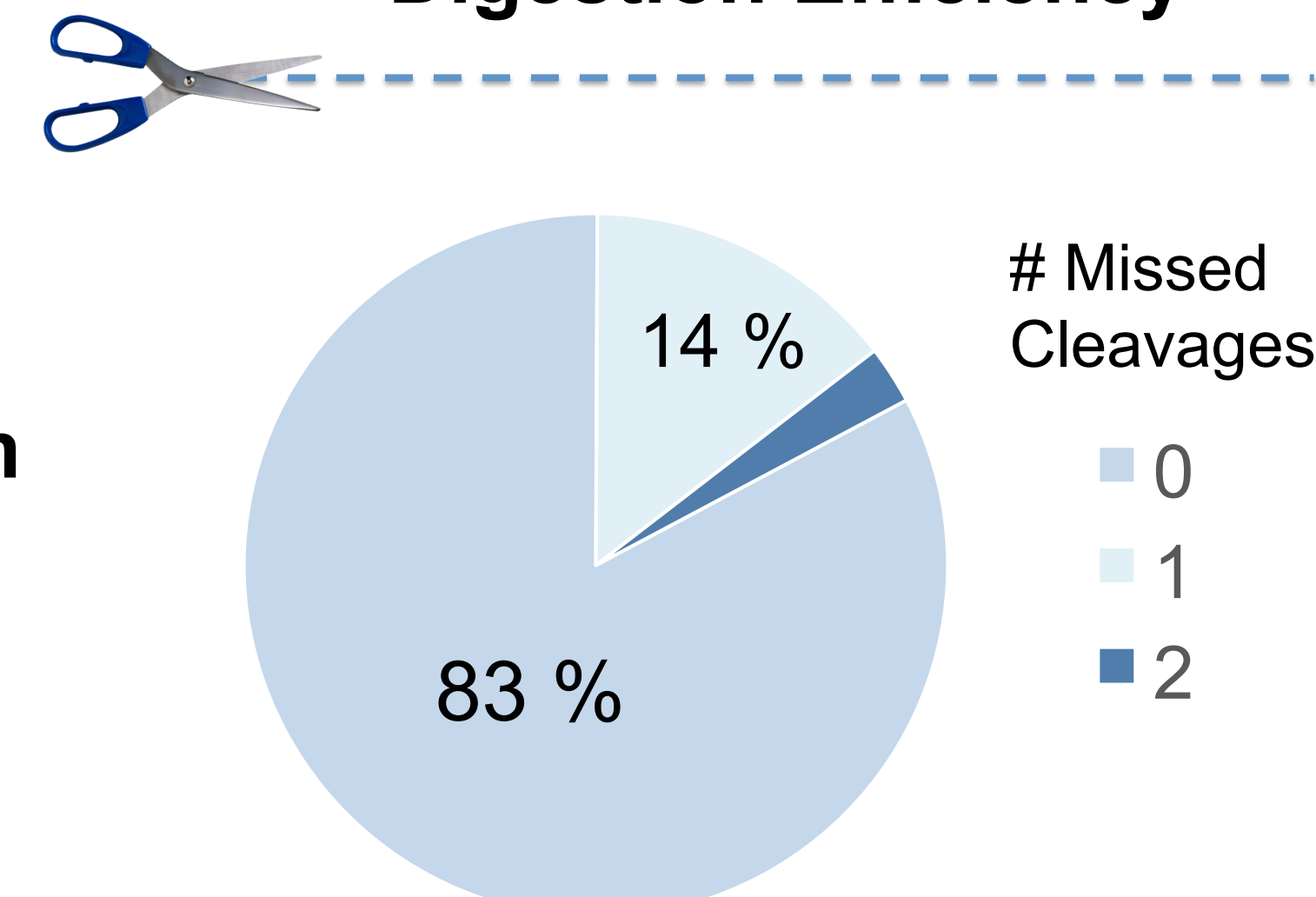


Bottom-up analysis of pellets and supernatants

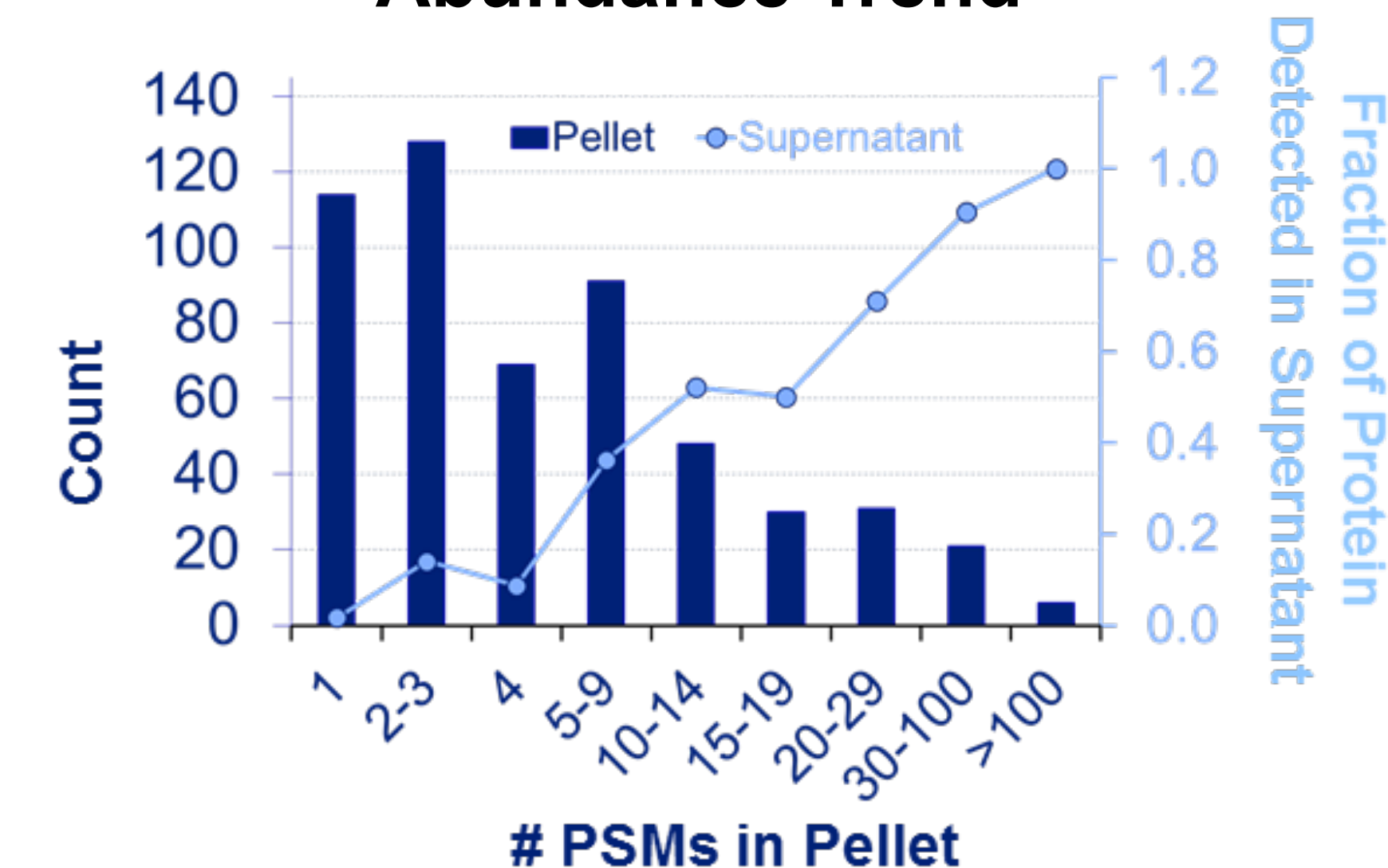
High Temperature → More Proteins



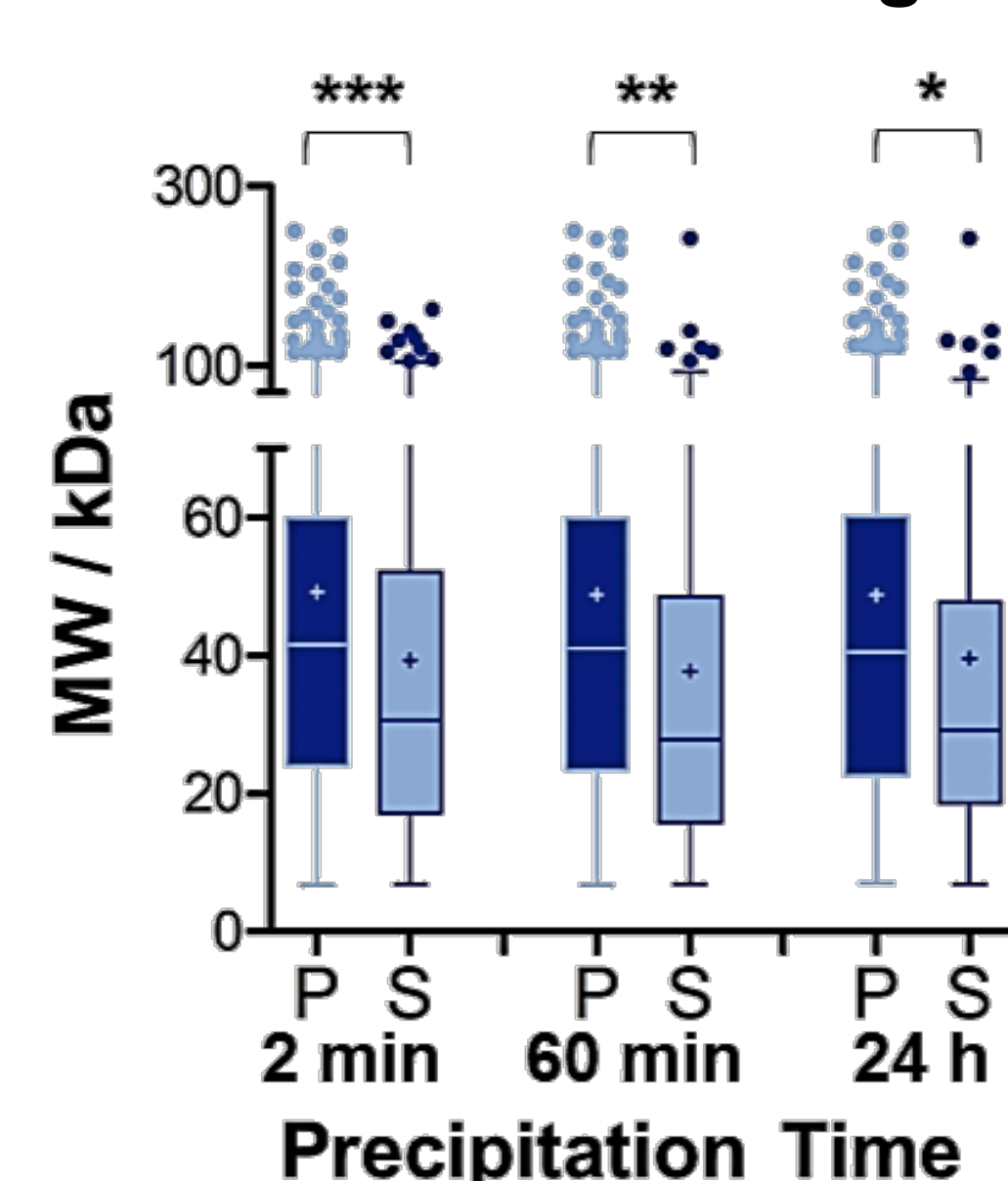
Digestion Efficiency



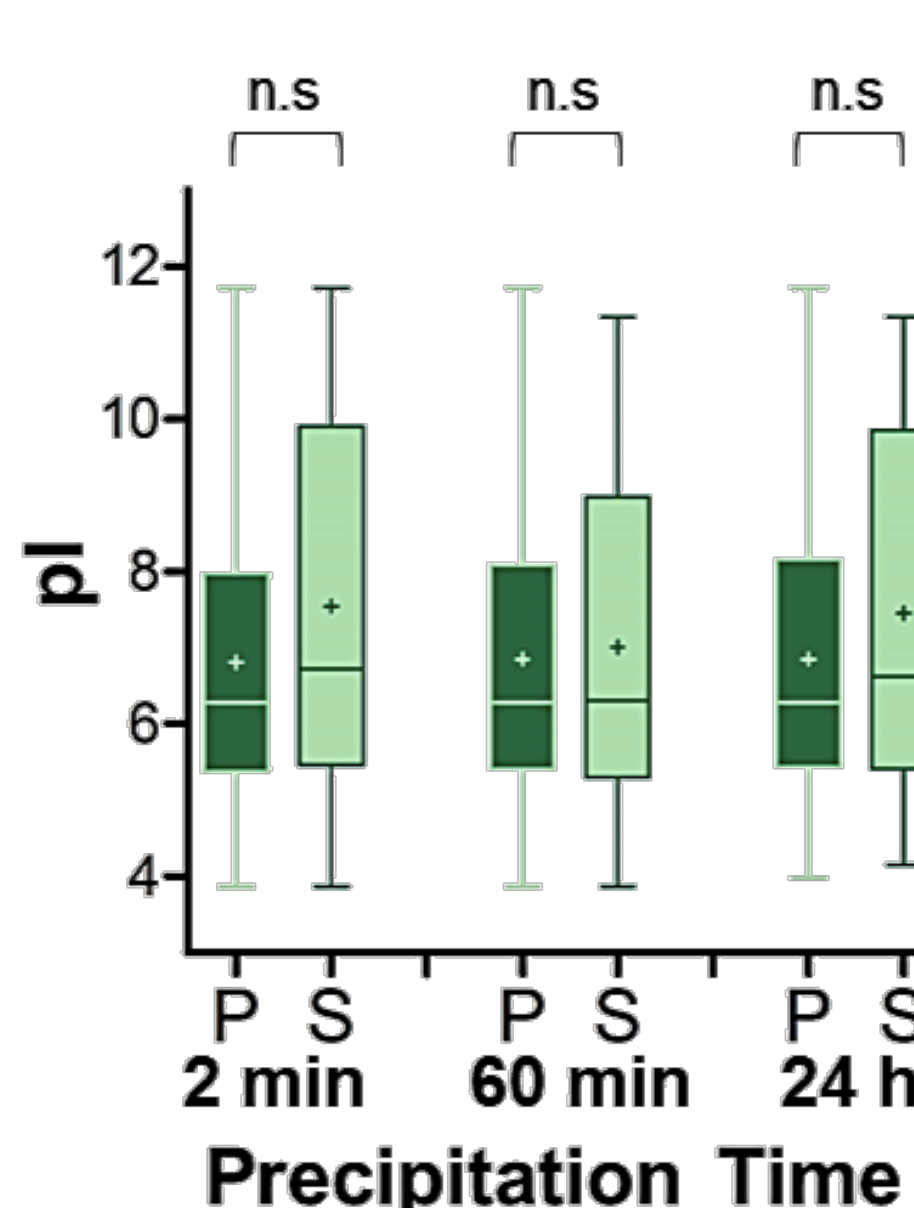
Abundance Trend



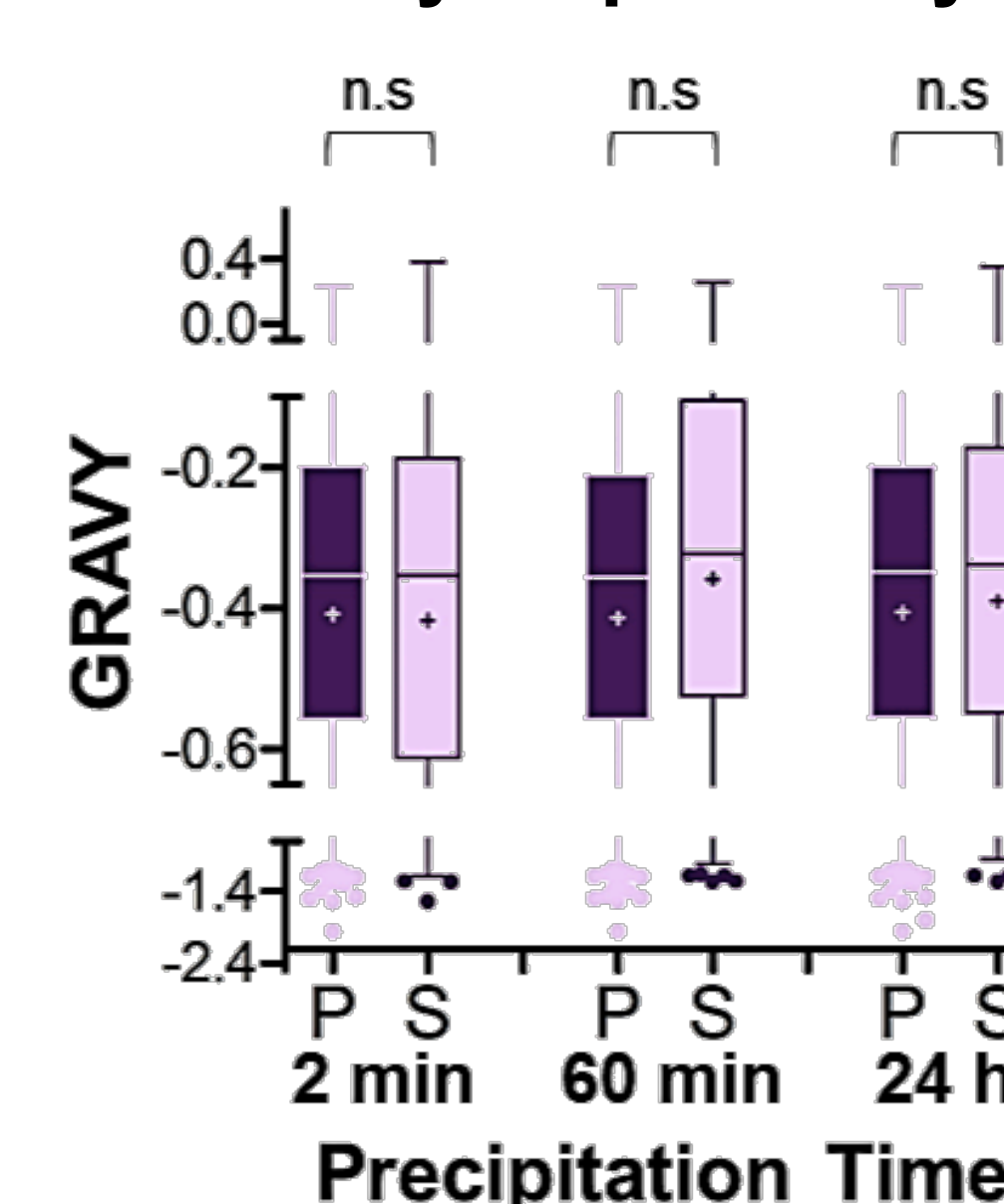
Molecular Weight



Isoelectric Point



Hydrophobicity



Sample Prep Efficiency of ProTrap XG

| | |
|---|--------------------|
| Recovery | 99 ± 1 % |
| Purity (SDS Depleted) | > 99.9 % |
| Digestion Efficiency (0 Missed Cleavages) | 82 % |
| Processing Time | 15 min + digestion |

CONCLUSIONS

- With >10 mM NaCl and 80% acetone, standard proteins and complex proteome mixtures can be quantitatively precipitated in 2 minutes at room temperature
- Dilute samples can be precipitated with equal efficiency to more concentrated samples
- The presence of a protein in the supernatant strongly correlates to its abundance. Most proteins identified in the supernatant are identified in the pellet with much greater abundance.
- There is no structural bias in which proteins are present in the pellet, though the recovery of low molecular weight proteins increases over time



Dalhousie's Chemistry Graduate Student Society

ACKNOWLEDGEMENTS

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REFERENCES

- Crowell, A., Wall, M., Doucette, A. (2013). Maximizing recovery of water-soluble proteins through acetone precipitation. *Analytica Chimica Acta*, 796, 48-54.
- Crowell, A., Maclellan, D., Doucette, A. (2015). A two-stage spin cartridge for integrated protein precipitation, digestion and SDS removal in a comparative bottom-up proteomics workflow. *Journal of Proteomics*, 118, 140-150.